A multifaceted peer reviewed journal in the field of Pharmacognosy and Natural Products www.phcog.com | www.phcog.net

Antioxidant, Acetylcholinesterase, Butyrylcholinesterase, and α-glucosidase Inhibitory Activities of *Corchorus depressus*

Samina Afzal, Bashir Ahmad Chaudhry, Ashfaq Ahmad¹, Muhammad Uzair, Khurram Afzal

Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Bahauddin Zakariya University, Multan, Pakistan, ¹Department of Pharmacology and Toxicology, School of Medicine, Virginia Commonwealth University, Richmond, USA

Submitted: 13-03-2017

Revised: 05-04-2017

Published: 13-11-2017

ABSTRACT

Background: Corchorus depressus (Cd) commonly known as Boa-phalee belonging to the family Tiliaceae having 50 genera and 450 species. Cd is not among the studied medicinal agent despite its potential in ethnopharmacology. Objectives: The present study investigated antioxidant, acetylcholinesterase (AChE), butyrylcholinesterase (BChE), and α -glucosidase inhibitory activities of Cd. The dichloromethane and methanolic extracts of the Cd were evaluated for biological activities such as antioxidant and enzyme inhibitory activities of AChE, BChE, and a-glucosidase. Materials and Methods: Antioxidant activity was evaluated by measuring free radical scavenging potential of Cd using 1,1-diphenyl-2-picrylhydrazyl. Enzyme inhibition activities were done by measuring optical density. Results: The methanol extract of roots of Cd showed potential free radical scavenging activity 99% at concentration 16.1 µg/ml. AChE was inhibited by aerial part of dichloromethane fraction by 46.07% \pm 0.45% while dichloromethane extracts of roots of Cd possessed significant activity against BChE with 86% inhibition compared with standard drug Eserine at concentration 0.5 mg/ml. The dichloromethane extract of roots of Cd showed 79% inhibition against α-glucosidase enzyme activity with IC₅₀62.8 \pm 1.5 µg/ml. **Conclusion:** These findings suggest Cd as useful therapeutic option as antioxidant and inhibition of AChE, BChE, and α-glucosidase activities.

Key words: 1,1-diphenyl-2-picrylhydrazyl, antioxidant, butyrylcholinesterase, dichloromethane, methanol

SUMMARY

- The aerial parts and roots of *Corchorus depressus* (*Cd*) were extracted in dichloromethane and methanol
- The extract of roots of *Cd* showed free radical scavenging activity 99% at concentration 16.1 µg/ml, Ach inhibition by aerial parts of dichloromethane fraction by 46.07%, and 79% inhibition against α -glucosidase enzyme activity with IC_{s0} 62.8 ± 1.5 µg/ml
- The dichloromethane and methanolic extracts of Cd exhibited antioxidant inhibition of acetyl cholinesterase, butyrylcholinesterase, and α -glucosidase activities.



Abbreviations used: DPPH: 1,1-diphenyl-2-picrylhydrazyl, *Cd: Corchorus depressus*, AChE: Acetylcholinesterase, BChE: Butyrylcholinesterase, AD: Alzheimer's disease.

Correspondence:

Dr. Samina Afzal,	
Department of Pharmaceutical Chemistry,	Access this article online
Faculty of Pharmacy, Bahauddin Zakariya University,	Website: www.phcog.com
Multan, Pakistan.	Quick Response Code:
E-mail: samina.afzal@bzu.edu.pk	TELC'S YARIES
Dr. Ashfaq Ahmad,	
Department of Pharmacology and Toxicology, Virginia	22392-273
Commonwealth University, Richmond, USA.	57.51.195i
E-mail: raza_chohan487@hotmail.com	
DOI: 10.4103/pm.pm_95_17	同時行移

INTRODUCTION

Nature at all times stands as a golden mark to demonstrate the exceptional phenomena of symbiosis. For a long time, natural products derived from plant and animal were used for the cure of diseases. In developing countries, approximately 80% of the people for their basic health care still depend on traditional medicine based on plant as well as animal species.^[11] The current demand and popularity of herbal medicines are increasing day by day. In ancient literature, approximately 500 plants are mentioned. In indigenous medicine system, about 800 plants have been used. Since ancient times, herbal medicines have been used in medical practices as the major remedy.

Antioxidants are nutrients in our foods which can avert or sluggish the oxidative damage to our body. Free radicals are also produced in different organs as result of metabolism of inhaled oxygen. Free radical damage may direct to cancer.^[2] Antioxidants act as free radical scavengers

and hence check and fix damage done by these free radicals. Now, the medicinal plants have become the aim for the hunt by cosmopolitan drug companies and study institutes for new drugs.

The inhibition of acetylcholinesterase (AChE) and butyrylcholinesterase (BChE), enzymes which breakdown acetylcholine (ACh) and butyrylcholine,

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

Cite this article as: Afzal S, Chaudhry BA, Ahmad A, Uzair M, Afzal K. Antioxidant, acetylcholinesterase, butyrylcholinesterase, and α -glucosidase inhibitory activities of *Corchorus depressus*. Phcog Mag 2017;13:647-51.

are thought as a promising approach for the management of Alzheimer's disease (AD).^[3] A potential source of AChE and BChE inhibitors is granted by the ample of plants in nature. AD is a progressive, neurodegenerative pathology that primarily affects the elderly population and is estimated to account for 50%-60% of dementia cases in persons over 65 years of age. In mammalian brain, there are two major forms of cholinesterases, namely, AChE and BChE. The most remarkable biochemical change in AD patients is a reduction of ACh levels in the hippocampus and cortex of the brain. Therefore, inhibition of AChE, the enzyme responsible for hydrolysis of ACh at the cholinergic synapse, is currently the most established approach to treating AD. While AChE is found in all excitable tissue, whether nerve or muscle, in most erythrocytes and in placental tissue, BChE is present more commonly in the body including within the central and peripheral nervous system, liver, and plasma.^[4] The serious side effects caused by licensed drugs used to treat AD have forced researchers to investigate safer AChE or BChE inhibitors from natural sources. Numerous plants and their constituents are reputed in traditional practices of medicine to enhance cognitive function and to alleviate other symptoms of AD, including depression.^[5]

In type 2 diabetes mellitus (DM), inhibition of α -glucosidase therapy is beneficial to delay absorption of glucose after a meal.^[6] α -glucosidase plays a role in the conversion of carbohydrates into glucose. By inhibiting α -glucosidase, glucose levels in the blood can be returned within normal limits.^[7] In spite of the introduction and extensive utilization of hypoglycemic agents, diabetes and the related complications continue to be a major health problem worldwide, which is affecting nearly 10% of the population worldwide and considered as a major cause of high economic loss which can in turn impede the development of nations. It is projected to become one of the world's main disablers and killers within the next 25 years.

Corchorus depressus (Cd) commonly known as Boa-phalee belonging to the family Tiliaceae. The Tiliaceae family has fifty genera and 450 species which are distributed in tropical and temperate regions, chiefly South Asia and South America. In Pakistan, about four genera and 24 species are found. The genus is enriched with pharmacological properties. Corchorus capsularis showed cardiovascular activity.^[8,9] Corchorus olitorius also showed spasmolytic activity,^[10,11] antihistaminic activity,^[12] hepatobiliary, renal and hematological activity,[13-15] antibacterial activity,[16] antiestrogenic, anticonvulsant activities,^[17] and antimalarial activity.^[18] Corchorus aestuans showed anticancerous activity.^[19] Cd showed analgesic and antipyretic activity.^[20,21] Literature survey of genus revealed phytochemical constituents triterpenoids, sterols,^[22] and flavonoids^[23] reported from chloroform extract of Cd. Cardiac glycosides,^[1] ionones glycosides, higher fatty acids, sterols, coumarins, and phenolics reported from leaves or seeds extraction of C. olitorius.[24-28] Various species of this genus have been used as folk medicine. Corchorus is used as a traditional medicine for the ailment of aches, dysentery, enteritis, fever, and tumors.^[29] The infusion of leaves is a demulcent, laxative, carminative, stimulant, appetizer, and tonic.^[30] The seeds of *Corchorus* are used for purgative, tonic, stomachic, fever, and in obstructions of abdominal problems.^[31] Jute fiber is obtained from C. capsularis and C. olitorius. It is used for treating respiratory treat infections and as a nervine and tonic; constituents include volatile oil, flavonoids, and phenolic acid.^[32]

Insight from literature review compelled us to investigate antioxidant and presence of possible AChE, BChE, and α glucosidase inhibitor activities of *Cd*.

EXPERIMENTAL PROCEDURES

Plant collection

The *Cd* was collected from Peruwal (District Khanewal) and identified by Professor Dr. Altaf Ahmed Dasti, plant Taxonomist, Institute of pure and applied biology, Bahauddin Zakariya University, Multan, Pakistan, whereas voucher specimen fl.p. 472/4 for *Cd* was deposited.

Extraction

Extraction method was followed as reported.^[33] The aerial parts and roots of *Cd* were cleaned, shade dried for 14 days, and pulverized to fine powdered in a mechanical grinder. The 1000 g of plant materials were subjected for extraction procedure using solvents dichloromethane and methanol at room temperature occasionally shaking for 24 h. Extracts were filtered by Buchner funnel. The filtrate extracts were concentrated by Rotavapor – R200 at 35°C. The dichloromethane and methanolic extracts of *Cd* were collected in separate sample bottles with designated different codes. The final extracts were obtained and used for *in vitro* antioxidant activity and AChE, BChE, and α glucosidase inhibitor activities. The results of the extraction along with the abbreviations used for different extracts are given in Table 1.

Phytochemical analysis

The dried and powdered aerial parts of Cd were investigated for the presence of alkaloids, anthraquinones, cardiac glycosides, tannins, and saponins as reported.^[33] The results of phytochemical analysis are given in Table 2.

Detection of alkaloids

Tem grams of the grinded plant material was boiled with 10 ml of acidified water in test tube for 1 min, cooled, and allowed the debris to settle. Filtered the supernatant liquid into another test tube. 1 ml of this filtrate was taken and 3 drops of Dragendorff's reagent were added; there was no precipitate. The remainder of filtrate was made alkaline by addition of dilute ammonia solution. A volume of 5 ml of chloroform was added to the solution in separating funnel; two layers were observed. The lower chloroform layer was pipetted out into another test tube. Chloroform layer was extracted by the addition of 10 ml of acetic acid and then discarded the chloroform. Then, extracts were divided into three portions, to one portion, few drops of Dragendorffs reagent were added and to the second portion, few drops of Mayer's reagent were added. Turbidity or precipitate was compared with the third untreated control portion.

Detection of anthraquinones glycosides

Two gram of powdered plant material was taken and extracted with 10 ml of hot water for 5 min, allowed it to cool and filtered; filtrate was extracted with 10 ml of carbon tetrachloride. Then, carbon tetrachloride layer was taken off, washed it with 5 ml water, and then, 5 ml dilute ammonia solution was added. No free anthraquinones were revealed as the absence of the appearance of pink to cherry red color in the ammoniacal layer.

Two gram of second sample of the plant was extracted with 10 ml of ferric

Table 1: Results	of the extraction	of the plant	Corchorus depressus
------------------	-------------------	--------------	---------------------

Plant name	Part used	Solvent	Weight of extract (g)	Abbreviation for the extracts
Cd	Aerial parts	Dichloromethane	39.85	CDAD
	(1000 g)	Methanol	7.95	CDAM
Cd	Roots	Dichloromethane	9.4	CDRD
	(1000 g)	Methanol	17.84	CDRM

Table 2:	Results o	f phytoo	hemica	l screening	of	Corc	horus c	lepressus
----------	-----------	----------	--------	-------------	----	------	---------	-----------

Name of plants	Alkaloid	Anthraquinone	Cardiac glycosides	Saponins
Cd	+	+	+	+
<i>Cd</i> (root part)	+	+	+	+

Cd: Corchorus depressus

chloride solution and 5 ml of hydrochloric acid; then, it was heated on water bath for 10 min and filtered. Filtrate was cooled and treated as above.

Detection of cardioactive glycosides

One gram of ground plant material was taken in a test tube, and 10 ml of 70% alcohol was added. It was then boiled for 2 min and filtered. Filtrate was diluted twice of its volume with water, and then, 1 ml of strong lead subacetate solution was added. This treatment leads to the precipitation of chlorophyll and other pigments, which were then filtered off. Filtrate was extracted with an equal volume of chloroform. Chloroform layer was pipetted out and evaporated to dryness in a dish over a water hath. Residue was dissolved in 3 ml of 3.5% ferric chloride in glacial acetic acid and was transferred to test tube after leaving for 1 min. A volume of 1.5 ml of sulfuric acid was then added, which formed a separate layer at the bottom. Cardioactive glycosides were revealed the appearance of brown color at interface (due to deoxy sugar) on standing and appearance of pale green color in the upper layer (due to the steroidal nucleus).

Detection of tannins

Prepare 10% w/v aqueous extract of grinded drug by boiling it with distilled water for about 10-20 min. Filtered the extract and performed the chemical tests with clear solution.

Ferric chloride test

A volume of 2 ml of ferric chloride solution was mixed to 1-2 ml clear solution of extract. A blueback precipitate indicated the presence of hydrolysable tannin.

Catechin test

Dip the match stick in plant extract, dry, and then, moist it with concentrated hydrochloric acid. Warm near flame, a red or pink wood is produced which shows the presence of catechin.

Determination of 1,1-diphenyl-2-picrylhydrazyl radical scavenging activity

The free radical scavenging activity was measured by 1,1-diphenyl-2-picrylhydrazyl (DPPH). The DPPH stock solution was prepared by dissolving 20 mg DPPH in 100 ml 95% methanol. This stock solution was stored at 20°C until needed not >10 days. DPPH working solution was prepared by diluting the stock of DPPH solution by adding methanol, and absorbance was adjusted about 0.980 \pm 0.02 at wavelength 517 nm using the spectrophotometer. A volume of 3 ml aliquot of this working solution mixed with 100 μ l of the plant samples at five different varying concentrations (4–322 μ g/ml). The solutions in the test tubes were shacked well and put in dark for 15 min at room temperature. Then, again the absorbance was measured at 517 nm. The percentage scavenging activity was determined based on the percentage of DPPH radical scavenged by using the following equation.^[34]

Scavenging effect (%) = {control absorbance – sample absorbance/ control absorbance} × 100.

Determination of acetylcholinesterase inhibitory activity

The evaluation of AChE inhibitory activity should be performed using thin layer chromatography (TLC) plates. These plates were treated with acetone. These plates were dried completely before their use. The solvents were removed from TLC plates completely by using hairdryer. Enzyme stock solution was then sprayed on the plates and again dried. The plates were put in flat position using plastic plugs in a tank with some water. The plates were covered and subjected to incubation at 37C for 20 min. The mixture containing 1-napththyl acetate (250 mg in 100 ml of ethyl alcohol) and Fast Blue B salt amounting to 400 mg in distilled water was prepared and used for detection of the enzyme. After incubation process, the mixture of the naphthyl acetate 10 ml and 40 ml of the Fast Blue B salt was prepared and sprayed on the plate so that there is purple color after a few minutes.^[35]

In vitro α -glucosidase inhibitory activity

A volume 135 μ l of 50 mM phosphate saline buffer pH (6.8) was dispensed in the 96 well plate. 20 μ l of the test sample in 70% dimethyl sulfoxide dispensed into the wells. 20 μ l of the enzyme was added into the wells and incubate the plate for 15 min. After incubation, preread of the plate was taken by the spectra max. After the preread, 25 μ l of the substrate was added and readings were taken on spectra max at 400 nm for 30 min. In the end, normal read is taken and the percent inhibition was calculated.

RESULTS AND DISCUSSION

Phytochemical screening showed the presence of alkaloids, anthraquinones, cardiac glycosides, tannins, and saponins as shown in Table 2.

In DPPH radical scavenging assay, the methanolic extract of roots of *Cd* showed maximum radical scavenging activity 99% at concentration 161 μ g/ml and 80% at concentration 16 μ g/ml. The methanolic extract of aerial parts of *Cd* showed 80% radical scavenging activity at high concentration 161 μ g/ml as shown in Table 3.

DPPH stable free radical method is a sensitive way to determine the antioxidant activity of plant extract. The methanolic extracts of the aerial and roots of the plant showed the strongest DPPH radical scavenging activity as compared to dichloromethane extracts of Cd as shown in Figure 1. This finding indicates that active constituents of plants are in methanolic fraction and mostly in root parts as compared to aerial parts of same fraction. The therapeutic potential of natural medicinal plants

Table 3: 1,1-diphenyl-2-picrylhydrazyl radical scavenging activity of different
extracts of Corchorus depressus

Plant extracts	Perce	IC ₅₀ (μg/ml)			
	161 µg/ml				
CDRD	23	10	5	2	Nil
CDRM	99	80	40	16	41.23±6.34
CDAM	80	30	18	10	20±6.60
CDAD	33	10	6	4	Nil
P control	97	93	89	56	

Ascorbic acid was used as standard



Figure 1: 1,1-diphenyl-2-picrylhydrazyl free radical scavenging activity of different fractions of different parts of *Corchorus depressus*

as an antioxidant in free radical-induced tissue injury, suggests that antioxidant activities of medicinal plant can be therapeutically useful for treatment.

IC50 values of different fractions and different parts of the plant Cd (concentration of the samples) required to inhibit the 50% population of the enzymes ACh esterase, and BChE were calculated as shown in Table 4. Cd did not show any promising activity against enzyme ACh esterase except aerial parts of dichloromethane fraction which showed $46.07\% \pm 0.45\%$ inhibition. At the same time, dichloromethane fraction of root part of Cd showed 86.36 \pm 0.65 inhibition of BChE with IC₅₀ value $132.8 \pm 0.87 \,\mu g$ as shown Table 4.

The result showed that dichloromethane extracts of aerial and roots of Cd possessed significant inhibition against ACh esterase and BChE inhibitory activities, respectively, as among other extracts compared with standard drug. Although it is indicating the effects of different fractions of Cd on CNS, these effects have not been observed in situation of enhanced cholinergic activity. These results are important because of the growing body of evidence suggesting that anticholinergic medications contribute to memory impairment in older adults.^[36-40] Cd like rivastigmine can be used as therapeutic option for the treatment of AD for having dual inhibitory effects on AChE and BChE esterase. Although Cd dominates against BChE esterase activity when compared to AChE, multifarious role can help to treat AD and problem associated with dementia. In case of AD, the ratio between BChE and AChE in the cortical region of brain changes from 0.5 to as high as 11.^[41] This indicates the supporting role of BChE in the regulation of AChE making functional importance of this enzyme in AD. Inhibiting both enzymes using Cd increased the ethnopharmacological importance of Cd.

The dichloromethane extract of roots of Cd showed 79% inhibition against α -glucosidase enzyme activity with IC₅₀ 62.8 ± 1.5 µg/ml while others are inactive as shown in Table 5. The glucosidase inhibition enzyme activity of dichloromethane extract of roots of Cd showed better responses when compared to standard drug with lower IC50 values. This indicates the potential use of Cd in diabetes can be a future therapeutic outcome. The α -glucosidase enzyme inhibitor has been used in DM.^[42,43]

Table 4: Results of acetylcholinesterase and butyrylcholinesterase inhibitory activities of Corchorus depressus

Code	AChE (%) at 0.5 mg/ml	AChE (IC ₅₀) ug/ml	BChE (%) at 0.5 mg/ml	BChE (IC ₅₀) ug/ml
CDAD	46.07±0.45	Nil	37.72±0.39	Nil
CDAM	23.93±0.63	Nil	60.49±0.89	Nil
CDRD	4.64±0.75	Nil	86.36±0.65	132.8±0.87 μg
CDRM	15.00 ± 0.85	Nil	67.72±0.99	
Control	Eserine	$0.04{\pm}0.001$	Eserine	$0.85 {\pm} 0.001$

AChE: Acetylcholinesterase; BChE: Butyrylcholinesterase

Table 5: Results of α-glucosidase against inhibition of dichloromethane and methanol extracts of Corchorus depressus

Sample		α -glucosidase			
code	Concentration (mg/ml)	Percentage inhibition	lC ₅₀ ±SEM (µg/ml)		
CDRD	0.5	79.2	62.8±1.5		
CDRM	0.5	44	Inactive		
	Standard dr	ug (control)			
Acarbose	0.64	59.1	83.33±0.34		
SEM: Standard error of mean					

A: Standard error of me

CONCLUSION

Aerial and root parts of methanolic and dichloromethane fractions of Cd possess antioxidant activity, AChE, BChE, and α -glucosidase inhibitory activities. However, the dichloromethane extract of roots showed better BChE and α -glucosidase inhibitory activities than the other fractions.

Acknowledgement

The project was supported by B. Z. University Multan, Pakistan. We also wish to acknowledge the technical support of H.E.J. Research Institute of Chemistry, International Center for Chemical and Biological Sciences, University of Karachi-75270, Karachi, Pakistan.

Financial support and sponsorship

The project was supported by B. Z. University Multan, Pakistan.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- 1. Proestos C. Lytoudi K. Mayromelanidou OK. Zoumpoulakis P. Sinanoglou VJ. Antioxidant capacity of selected plant extracts and their essential oils. Antioxidants (Basel) 2013;2:11-22.
- 2. Sultana B, Hussain Z, Hameed M, Mushtaq M. Antioxidant activity among different parts of aubergine (Solanum melongena L.). Pak J Bot 2013;45:1443-8.
- 3. Talić S, Dragičević I, Ćorajević L, Martinović Bevanda A. Acetylcholinesterase and butyrylcholinesterase inhibitory activity of extracts from medicinal plants. Bull Chem Technol Bosnia Herzegovina 2014:43:11-4.
- 4. Orhan I, Kartal M, Naz Q, Ejaz A, Yilmaz G, Kan Y, et al. Antioxdant and anticholinesterase evaluation of selected Turkish Salvia species. Food Chem 2007:103:1247-54.
- 5. Politeo O, Botica I, Bilusić T, Jukić M, Carey I, Burcul F, et al, Chemical composition and evaluation of acetylcholinesterase inhibition and antioxidant activity of essential oil from Dalmatian endemic species Pinusnigra Arnold ssp. Dalmatica (Vis.) Franco. J Med Plants Res 2011:5:6590-6.
- 6. Kim KY, Nam KA, Kurihara H, Kim SM, Potent alpha-alucosidase inhibitors purified from the red alga Grateloupia elliptica. Phytochemistry 2008;69:2820-5.
- 7. Bosenberg LH, Van Zyl DG. The mechanism of action of oral antidiabetic drugs: A review of recent literature . J Endocrinol Metab Diabetes S Afr 2008:13:80-8
- 8. Karrer P, Banerjee P. Corchortoxin, a cardiac agent from jute seeds. Helv Chim Acta 1949:32:2385-92
- 9. Frerejacque M, Durgeat M. Digitalis like poisons of jute seed. Compt Rend 1954;238:507-9.
- 10. Khatib S, Alkofahi A, Hasan M, Najib N, El-Oglah A. Effect of Corchorus olitorius seed extract on left ventricular pressure, coronary flow and heart rate of the isolated rabbit heart. Biomed Lett 1998;57:76-83
- 11. Leshchinskii LA, Karbasnikova GV, Zabel'ian OM, Vasil'kova AA. Use of cardiac glycosides of strophanthin-like action in the pharmacotherapy of chronic circulatory insufficiency Kardiologija 1978:18:76-83.
- 12. Yoshikawa M, Murakami T, Shimada H, Yoshizumi S, Saka M, Yamahara J, et al. Medicinal food stuffs XIV, on the bioactive constituents of Moroheiya (2): New fatty acids, corchorifatty acids A. C. D. E and E from the leaves of Corchorus olitorius L (Tiliaceae); Structures and inhibitory effect on NO production in mouse peritoneal macrophages. Chem Pharm Bull 1998;46:1008-14.
- 13. Laskar S, Majumdar SG, Basak B, Dey CD. Influence of jute (Corchorus olitorius) seed protein enriched diet on some enzymes and liver lipids of albino rats (Rattus norvegicus). Physiol Bohemoslov 1986:35:86-9.
- 14. Inami S, Tabata K, Shimizu J, Kusunoki K, Ishida H, Matsuguma M, et al. Dried green leaf powders of Jew's mellow (Corchorus olitorius), persimmon (Diosphyros kaki) and sweet potato (Ipomoea batatas Poir) lower hepatic cholesterol concentration and increase fecal bile acid excretion in rats fed a cholesterol-free diet. Plant Foods Hum Nutr 1998;52:55-65.
- 15. Mazumder U. Gupta M. Pal D. Bhattacharva S. Chemical and toxicological evaluation of methanol extract of Cuscuta reflexa Roxb stem and Corchorus olitorius Linn seed on haematological parameters and hepatorenal functions in mice. Acta Pol Pharm 2003;60:317-23.

- Ibrahim T, Fagbohun E. Physicochemical properties and *in vitro* anti- bacterial activity of Corchorus olitorius Linn. Seed oil. Life Sci Leafl 2011;15:499-505.
- Gupta M, Mazumder U, Pal D, Bhattacharya S. Antisteroidogenic activity of methanolic extract of *Cuscuta reflexa* Roxb stem and *Corchorus olitorius* Linn seed in mouse ovary. Indian J Exp Biol 2003;4:641-4.
- Sathiamoorthy P, Lugasi E, Schlesinger P, Kedar I, Gopas J, Pollack Y, et al. Screening for cytotoxic and anti-malarial activities in desert plants of the Negev and Bedouin market plant products. Pharm Biol 1999;37:188-95.
- Bhakuni D, Dhar M, Dhar M, Dhawan B, Mehrotra B. Screening of Indian plants for biological activity part II. Indian J Exp Biol 1969;7:250-62.
- Vohora SB, Shamsi MA, Khan MS. Antipyretic and analgesic studies on the diacetate of a new triterpenic acid isolated from *Corchorus depressus* L. J Ethnopharmacol 1981;4:223-8.
- Ikram M, Khattak S, Gilani S. Antipyretic studies on some indigenous Pakistani medicinal plants II. J Ethnopharmacol 1987;19:185-92.
- Ahmad V, Ali A, Ali Z, Baqai F, Zafar F. Cycloartane triterpene glycosides from *Corchorus depressus*. Phytochemistry 1998;49:829-34.
- Harsh M, Nag T. Flavonoids with antimicrobial activities of arid zone plants. Geobios 1988;15:32-35.
- Mahato S, Pal B, Sarkar S. New triterpenoids saponins from *Corchorus acutangulus*. Phytochemistry 1988;27:1433-8.
- 25. Yoshikawa M, Shimada H, Saka M, Yoshizumi S, Yamahara J, Matsuda H. Medicinal foodstuffs. V. Moroheiya (1): Absolute stereo structures of corchoionosides A, B and C, histamine inhibitors from the leaves of Vietnamese *Corchorus olitorius L*. (Tiliaceae). Chem Pharm Bull 1997;45:464-9.
- Manzoor-I-Khuda M, Islam A. Chemical constituents of *Corchorus olitorius* and *Corchorus capsularis* (jute).II. isolation of corosin & sitosterol from roots. Pak J Sci Ind Res 1971;14:49-56.
- Mukherjee K, Mitra S, Ganguly S. A new coumarin from the seeds of jute (*Corchorus olitorius L.*). Nat Prod Sci 1998;4:51-2.
- Azuma K, Nakayama K, Koshioka M, Ippoushi K, Yamaguchi Y, Kohata K, et al. Phenolic antioxidants from the leaves of *Corchorus olitorius L*. J Agric Food Chem 1999;47:3963-6.
- Duke J, Wain K. Medicinal plants of the world. Computer index with more than 85,000 entries. Vol. 3. Agriculture Research Service, Beltsville, Maryland; 1981.
- 30. Kritikar K, Basu B. Indian Medicinal Plants. Lalit Mohan Basu, Allahabad; 1935. p. 402-3.

- Chopra R, Nayar S, Chopra I. Glossary of Indian Medicinal Plants. New Delhi, India: Council of Scientific and Industrial Research; 1986.
- Evans WC. Trease and Evans Pharmacognosy. 15th ed. New York, U.S.A: Saunders/Elsevier; 2002. p. 20-9.
- Afzal S, Chaudhary BA, Ahmad A, Afzali K. Prelimiary phytochemical analysis and antifungal activities of crude extracts of *Zaleya pentandra* and *Corchorus depressus* Linn. Acta Pol Pharm 2015;72:329-34.
- Cuendet M, Hostettmann K, Potterat O. Iridoid Glucosides with free radical scavenging properties from *Fagraea blumei*. Helv Chim Acta 1997;80:1144-52.
- Marston A, Kissling J, Hostettmann K. A rapid TLC bio autographic method for the detection of Acetyl cholinestrase and butyryl cholinestrase inhibitors in plants. Phytochem Anal 2002;13:51-4.
- Mulsant BH, Pollock BG, Kirshner M, Shen C, Dodge H, Ganguli M. Serum anticholinergic activity in a community-based sample of older adults: Relationship with cognitive performance. Arch Gen Psychiatry 2003;60:198-203.
- Han L, McCusker J, Cole M, Abrahamowicz M, Primeau F, Elie M. Use of medications with anticholinergic effect predicts clinical severity of delirium symptoms in older medical inpatients. Arch Intern Med 2001;161:1099-105.
- Ancelin ML, Artero S, Portet F, Dupuy AM, Touchon J, Ritchie K. Non-degenerative mild cognitive impairment in elderly people and use of anticholinergic drugs: Longitudinal cohort study. BMJ 2006;332:455-9.
- Flacker JM, Cummings V, Mach JR Jr., Bettin K, Kiely DK, Wei J. The association of serum anticholinergic activity with delirium in elderly medical patients. Am J Geriatr Psychiatry 1998;6:31-41.
- Lechevallier-Michel N, Molimard M, Dartigues JF. Drugs with anti-cholinergic properties and cognitive performance in the elderly: Results from the PAQUID study. Br J Clin Pharmacol 2005;59:143-51.
- Perry EK, Tomlinson BE, Blessed G, Bergmann K, Gibson PH, Perry RH. Correlation of cholinergic abnormalities with senile plaques and mental test scores in senile dementia. Br Med J 1978;2:1457-9.
- 42. Bati K, Kwape TE, Chaturvedi P. Anti-diabetic effects of an ethanol extract of *Cassia abbreviata* stem bark on diabetic rats and possible mechanism of its action: Anti-diabetic properties of *Cassia abbreviata*. J Pharmacopuncture 2017;20:45-51.
- 43. Giordani MA, Collicchio TC, Ascêncio SD, Martins DT, Balogun SO, Bieski IG, et al. Hydroethanolic extract of the inner stem bark of *Cedrela odorata* has low toxicity and reduces hyperglycemia induced by an overload of sucrose and glucose. J Ethnopharmacol 2015;162:352-61.