















## RESULTS AND DISCUSSION

### Morphological differences between *Cyperus scariosus* and *Cyperus rotundus*

In CS, the stems are slender, three-sided and triangular in cross-section. An umbrella-like tuft of long narrow leaves occurs at the top. Leaves are whorled, lanceolate and green in color, with a distinct ridge. The rhizomes are initially white in color and eventually turn brown with growing age. Lateral shoots arise from the base of the stem in an immediately ascending manner. Whereas flowers are initiated from axillary buds. In contrast, CR is a grass-like weed with an extensive underground network of basal bulbs, fibrous roots, thin, wiry rhizomes and tubers born in chains of 2–6 or more on rhizomes. The leaves are mostly basal, dark green, with a prominent midrib and an abrupt taper at the top. The purplish to red-brown inflorescence is born on a stem that is triangular in cross-section and usually taller than the foliage [Figure 1].

### Comparative phytochemical analysis between *Cyperus scariosus* and *Cyperus rotundus*

The rhizome powders showed a distinct color variation between the species [Figure 2 (I. *Cyperus Scariosus* and II. *Cyperus rotundus*)]. However, classifying the species based on color can be erratic and misleading. Based on the preliminary phytochemical analysis of various solvent extracts of CS and CR constituents of terpenoids and steroids were found in excess amount, constituents such as alkaloids, glycosides, carbohydrates, phenols, fats, and oils were also found. The compounds related to tannins; saponins and flavonoids were found to be absent [Table 2]. In TLC analysis hexane extracts of both the plant species showed better separation in hexane: EtOAc (90:10), whereas chloroform extracts showed better separation in chloroform: Acetone (70:30) as mobile phase. In case of methanolic extract in CS showed separation in  $\text{CHCl}_3$ : MeOH (60:40), whereas in case of CR methanolic extract (40:60)  $\text{CHCl}_3$ : MeOH as the mobile phase [Figure 2 (Ic and Iic)]. Based on these TLC profile pattern and their retardation factor (Rf), it is suggested that both the plant species as different.

Further, LC–tandem mass spectrometry and GC/MS techniques were used to determine the chemical profiles of CR and CS. Thus, liquid chromatogram patterns of CS hexane extract showed major peaks at their retention times 2.89, 3.359, 9.363, and 10.84 [Figure 3a-c]. Whereas in case of CS chloroform extract major peaks were observed at retention times 3.128, 4.99, 8.12 and 9.210 [Figure 4a-c]. Similarly, hexane extract of CR displayed major peaks at 3.12, 4.2, 7.81, 9.11, and 9.82 [Figure 5a-c], whereas in chloroform extract of CR major peaks observed at retention times 2.686, 3.84, 4.26, 5.82, 7.90, and 9.9, respectively [Figure 6a-c]. Furthermore, comparisons of the mass spectral patterns of hexane and chloroform extracts of CS indicated a molecular ion peaks at retention times of 3.3 and 3.1 min with  $m/z$  415.1  $[\text{M}^+]^+$  [Figures 3d and 4d], which was correlated to the  $\beta$ -sitosterol from the literature study.<sup>[11]</sup> In contrary, the mass spectra of hexane and chloroform extracts of CR showed the ion peak at  $m/z$  219.1  $[\text{M}^+]^+$  with retention time 7.8 min correlated to the compounds  $\alpha$ -cyperone and cyperotundone from the literature data.<sup>[12]</sup> In addition to this the ion peak at 26.3 min retention times with  $m/z$  216.1  $[\text{M}^+]^+$  [Figures 5d and 6d] correlated to the compounds  $\alpha$ -cyperene and Isocyperol<sup>[13]</sup> in both the extracts of CR. Further, the ion peak with  $m/z$  413.2  $[\text{M}^+]^+$  at retention time 30.3 min correlated to stigmasterol<sup>[14]</sup> and the ion peak at 6.3 min with  $m/z$  427.2  $[\text{M}^+]^+$  [Figure 6d] correlated to the Lupeol in chloroform extract of CR from the literature data.<sup>[15]</sup>

GC-MS chromatograms of hexane extract from the rhizomes of CS showed 30 peaks and chloroform extract showed 22 peaks. Whereas CR hexane extract showed, 23 peaks and chloroform extract showed 15 peaks. These chromatograms with retention time were shown

in Figure 7. By comparing GC-MS spectra of with NIST library, we identified common and unique compounds between these extracts in the form of Venn diagram [Figure 8]. This diagram depicts the common compounds presented in CS-hexane and CS- $\text{CHCl}_3$  extracts were seven. Five compounds were common in CS- $\text{CHCl}_3$  and CR-hexane. In CS-hexane, CS- $\text{CHCl}_3$  and CR-hexane two compounds were similar. Five compounds were similar between CR-hexane and CR- $\text{CHCl}_3$ . One compound is similar in CS- $\text{CHCl}_3$ , CR-hexane, and CR- $\text{CHCl}_3$  between these extracts. In CS-hexane, CS- $\text{CHCl}_3$ , and CR- $\text{CHCl}_3$  one compound is similar. Finally, in CS-hexane and CR- $\text{CHCl}_3$  one compound is similar. In contrast to unique compounds, 27 compounds were unique in CS-hexane, 14 compounds in CS- $\text{CHCl}_3$ , 15 compounds in CS-hexane and 13 compounds in CR- $\text{CHCl}_3$ . The name of the identified compounds, molecular weight, and their molecular formula were presented in Tables 3 and 4.

### Isolation and structural elucidation of compounds in *Cyperus scariosus*

The concentrated hexane and chloroform extracts from the rhizomes of were chromatographed on silica gel and the resultant fractions and repeated column chromatography purification of resultant fractions led to the isolation of three compounds. The structures of isolates were established using IR, MS, 1D, and 2D NMR spectroscopic techniques. After comparing their spectral data with those reported in the literature<sup>[16]</sup> they were identified as known compounds and confirmed as stigmasterol,  $\beta$ -sitosterol and lupeol [Figure 9]. These compounds were found to be major constituents in both the species that is, CR<sup>[17]</sup> and CS.

## CONCLUSION

In this study, we examined the morphological and chemoprofiling pattern of CS and CR to systematically classify these species. Based on their morphological attributes, it is found and confirmed that these two species are different. Chemoprofiling analyses revealed some of the common phytochemical compounds similar in between these herbs. Finally, we conclude that these two herbs hold some of the similar phytochemical compounds in major quantity but are morphologically different.

### Acknowledgments

Authors would like to thank the management of KLEF University and Director, CSIR-Indian Institute of Chemical Technology for providing facilities and also thankful to Mr. Pardhasaradhi Mathi for his technical support. One of the authors, Lavanya Kakarla is thankful to DST, New Delhi for financial support under Women scientist (WOS-A) scheme.

### Financial support and sponsorship

This work is financially supported by Department of Science and Technology, New Delhi, India, under the Women Scientist (WOS-A) project grant (SR/ WOS-A/ LS-1160/ 2014 (G).

### Conflicts of interest

There are no conflicts of interest.

## REFERENCES

- Rajkumar V, Guha G, Kumar RA. Isolation and bioactivity evaluation of two metabolites from the methanolic extract of *Oroxylum indicum* stem bark. *Asian Pac J Trop Biomed* 2012;2:S7-11.
- Dahanukar S, Kulkarni R, Rege N. Pharmacology of medicinal plants and natural products. *Indian J Pharmacol* 2000;32:S81-8.
- Cragg GM, Newman DJ. Natural products: A continuing source of novel drug leads. *Biochim Biophys Acta* 2013;1830:3670-95.
- Penza M, Montani C, Jeremic M, Mazzoleni G, Hsiao WL, Marra M, et al. MAK-4



- and -5 supplemented diet inhibits liver carcinogenesis in mice. BMC Complement Altern Med 2007;7:19.
5. Wadhwa M, Bakshi M. Herbal feed additives impact on the rumen environment in buffaloes. Indian J Anim Nutr 2006;23:102-9.
  6. Kakarla L, Mathi P, Allu PR, Rama C, Botlagunta M. Identification of human cyclooxygenase-2 inhibitors from *Cyperus scariosus* (R.Br) rhizomes. Bioinformation 2014;10:637-46.
  7. Mitra S, Saxena E, Dixit M. Natural Composition for Curing Hepatitis-B, Methods for Making the Same and Pharmaceutical Formulations Thereof. In: Google Patents; 2005.
  8. Bhawna K, Kumar SS, Lalit S, Sharmista M, Tanuja S. *Cyperus scariosus*: A Potential medicinal herb. Int Res J Pharm 2013;4:17-20.
  9. Srivastava R, Singh A, Srivastava G, Lehri A, Niranjana A, Tewari S, et al. Chemical constituents and biological activities of promising aromatic plant *Nagarmotha* (*Cyperus scariosus* R. Br.): A review. Proc Indian Natl Sci Acad 2014;80:525-36.
  10. Yisa J. Phytochemical analysis and antimicrobial activity of *Scoparia dulcis* and *Nymphaea lotus*. Aust J Basic Appl Sci 2009;3:3975-9.
  11. Jaiswal Y, Liang Z, Guo P, Ho HM, Chen H, Zhao Z. Tissue-specific metabolite profiling of *Cyperus rotundus* L. rhizomes and (+)-nootkatone quantitation by laser microdissection, ultra-high-performance liquid chromatography-quadrupole time-of-flight mass spectrometry, and gas chromatography-mass spectrometry techniques. J Agric Food Chem 2014;62:7302-16.
  12. Sonwa MM, König WA. Chemical study of the essential oil of *Cyperus rotundus*. Phytochemistry 2001;58:799-810.
  13. Kim SJ, Kim HJ, Kim HJ, Jang YP, Oh MS, Jang DS. New patchoulane type sesquiterpenes from the rhizomes of *Cyperus rotundus*. Bull Korean Chem Soc 2012;33:3115.
  14. Sayed HM, Mohamed MH, Farag SF, Mohamed GA, Proksch P. A new steroid glycoside and furochromones from *Cyperus rotundus* L. Nat Prod Res 2007;21:343-50.
  15. Prakash CV, Prakash I. Isolation and structural characterization of lupane triterpenes from *Polypodium vulgare*. Res J Pharm Sci 2012;1:23-7.
  16. Deore SL, Khadabadi SS. Isolation and characterization of phytoconstituents from *Chlorophytum borivilianum*. Pharmacognosy Res 2010;2:343-9.
  17. Puratchikody A, Devi CN, Nagalakshmi G. Wound healing activity of *Cyperus rotundus* linn. Indian J Pharm Sci 2006;68:97-101.



Dr. K. Suresh Babu



Dr. Mahendran Botlagunta



Lavanya Kakarla

## ABOUT AUTHORS

**Dr. K. Suresh Babu**, is a scientist at the Natural Products Laboratory, Indian Institute of Chemical Technology (IICT), Hyderabad. His research interest are new drug discovery from natural products, structure-activity relationship studies and standardization of herbal drugs of commercial importance.

**Dr. Mahendran Botlagunta**, is a Associate professor from K L E F University, Guntur, Andhra Pradesh and recipient of UGC-Research award from UGC, New Delhi. His research focuses on interdisciplinary areas of cancer, Inflammation, nanomedicine and bioinformatics.

**Lavanya Kakarla**, is a Research scholar in Department of Biotechnology at K L E F University from 2011. She received Master's degree in the year 2007 from Bharathidasan University, Trichy. Her research interests include phytomedicine and Plant tissue culture. She has been awarded Women Scientist (WOS-A) in 2015 by DST, Government of India.