Curcumin attenuates neurotoxicity induced by fluoride: An in vivo evidence

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Submitted: 29-01-2013 Revised: 08-05-2013 Published: 07-02-2014

ABSTRACT

Background: Curcumin (Cur), an active ingredient of turmeric is known to have multiple activities, including an antioxidant property and has been suggested to be useful in treatment of several neurological diseases. Objective: To investigate the neuroprotective effects of Cur to mitigate the effect of the Fluoride (F) induced neurotoxicity in mice brain using the histological and the biochemical parameters. Materials and Methods: Exposure of mice (30 days old male) to F (120 ppm) daily for 30 days. Result and Discussion: Treatment with the F causes an increase in lipid peroxidation (LPO) and also increase in the neurodegenerative cells in the hippocampal sub-regions. Interestingly, co-treatment with Cur (30 mg/kg BW) with F (120 ppm) for 30 days results in significant decreases in LPO with a concomitant decrease in neurodegeneration as compared with those treated with F alone. Conclusion: Our study reveals that Cur is useful in ameliorating degenerative effects of F in mice brain.

Key words: Curcumin, neurodegeneration, neuroprotection, lipid peroxidation, oxidative stress, fluoride, histology.

INTRODUCTION

Fluoride (F) is probably the first inorganic ion which drew attention of the scientific world for its toxic effects and now the F toxicity through drinking water is well-recognized as a global problem. Health effect reports on F exposure also include various cancers, adverse reproductive activities, cardiovascular, and neurological diseases.[1,2] Major cause of F induced neurological diseases is excitotoxicity causing degeneration of neuron cell bodies in selective brain areas.[3,4] The process of neurodegeneration begins at the cellular level in which two main processes, oxidative stress, and excitotoxicity act relentlessly to inflict the majority of cell damage and death. Oxidative Stress can cause cellular damage and subsequent cell death mainly by apoptosis in neurodegeneration because the reactive oxygen species (ROS) oxidize vital cellular components such as lipids, proteins, and deoxyribonucleic acid;[6,8] therefore, the imbalance between intracellular ROS and antioxidant defense mechanisms results in oxidative stress that is harmful for neurons. F had obvious influence on phospholipid and fatty acid composition in brain cells of rats where lipid peroxidation (LPO) due to oxidative stress causes a decrease in total brain phospholipid content.[9] Thus, it is not surprising that oxidative stress is a common discussion point for neurodegenerative changes observed in fluorosis patients. Damage to neurons can reflect both an increase in oxidative processes and a decrease in antioxidant defenses. One proposed strategy for the treatment of neurodegenerative disorders that has gained considerable attention is the use of natural antioxidant agents since one common advantage of all these compounds is their human safety.[7] Curcumin (Cur) is an ancient Indian herb, which is extracted from the curry powder and has powerful antioxidant and anti-inflammatory properties.[10] It reacts with the free radical and can protect cell from singlet oxygen, hydroxyl radical, and superoxide radical damage. The increase in level of glutathione by Cur indicates its antioxidant property possibly by increasing the endogenous defense of the brain to combat oxidative stress.[11] Cur's neuroprotective role has recently been demonstrated during neuronal developmental[12] and adult hippocampal neurogenesis in fluoride intoxicated mice[13] and a biological activity that may enhance neuronal plasticity and repair.[13] Here, we investigated the possible protective effects of Cur, a herbal polyphenolic compound, with respect to F induced oxidative stress in hippocampal regions of mice brain.
**MATERIALS AND METHODS**

Drug and chemicals
Cur used in this study was purchased from Prolab marketing Ltd. All other chemicals used were purchased from the standard commercial suppliers and were of analytical grade.

Animals
Adult (one and half months), male, *Swiss Albino* mice weighing 30 g ± 5 g (BW) were used. Animals were divided into control and three experimental groups (I to III: *n* = 7). All experiments and protocols were approved by Institutional Animal Ethical Committee (IAEC Reg. No. 973/ac/06/CPCSEA). Animals were maintained in the laboratory as per international norms.

Control group
Group I: Control animals were given mice chow and F-free water ad-libitum for 30 days.

Experimental groups
Mice were randomly divided into three experimental groups and the dosages were selected on the basis of pilot studies conducted in our laboratory.

The treatments were given for 30 days to all the groups and the dosage are as follows:

<table>
<thead>
<tr>
<th>Experimental group</th>
<th>Doses</th>
<th>Mode of treatment</th>
</tr>
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<tbody>
<tr>
<td>II</td>
<td>Fluoride (120 ppm)</td>
<td>Fluoride was given in distilled drinking water ad libitum</td>
</tr>
<tr>
<td>III</td>
<td>Fluoride+Cur (120 ppm+30 mg/kg BW)</td>
<td>Oral dose of Cur dissolved in olive oil along with fluoride in drinking water</td>
</tr>
<tr>
<td>IV</td>
<td>Cur (30 mg/kg BW)</td>
<td>Oral dose of Cur dissolved in olive oil</td>
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Malondialdehyde (MDA) biochemical estimation
MDA content in brain tissue was measured by using the protocol of Buege and Aust, 1978. After 30 days, all the mice were weighed and sacrificed by cervical dislocation. The brain was dissected out and placed on chilled glass plate, dried, and weighed. Brains were cut into two sagittal pieces with the help of surgical blade, and the hippocampus was dissected out under a stereomicroscope. The tissue was minced, and then homogenized in chilled 100 mM pH-7.2 phosphate buffer saline using a Teflon mechanical homogenizer. Samples were then diluted tenfold and the homogenate was spun at 10,000 rpm for 15 min and the supernatant was used for enzymatic assay. To 0.8 ml of supernatant, 1.2 ml of TCA-TBA-HCl (Trichloroacetic acid (TCA): 15% *w/v*, thiobarbituric acid (TBA) 0.375%, hydrochloric acid (HCl) 0.25 N) reagent was added and kept in boiling water bath for 10 min. After cooling 2.0 ml of freshly prepared 1 N NaOH was added. The absorbance of pink color obtained was measured at 535 nm against blank, which contained distilled water.

Calculation
Molar extinction of MDA at 535 nm is 1.56 × 10^5/M/cm

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V \times \text{OD}_{535}/0.156 = 4 \times \text{OD}_{535} = 25.6 \times \text{OD}_{535} \text{ nmoles/ml of homogenate}
\]

Statistical analysis
The data were expressed as Mean ± SEM and comparisons of all groups were carried out using the ANOVA (one-way analysis) followed by turkey’s post test.

Data were analyzed by the prism software program (Graph pad software Inc.).

Cresyl violet studies
Mice were anesthetized with ether vapors and perfused transcardially using a perfusion unit. The brains were dissected and overnight post fixation was carried out in 10% formaldehyde. Post fixed brains were washed in 0.01 M Phosphate buffer (pH-7.4), three changes of 30 min each. The brain was then dehydrated in graded series of alcohol i.e., 30%, 50%, 70%, 90% and absolute alcohol. This was followed by clearing in xylene. The brain was then impregnated with 2 changes of 50% paraffin in xylene in an oven set at 59.5°C, followed by two changes in melted paraffin at 59.5°C. The blocks were prepared with the help of Leukhart’s pieces and tissues were oriented to cut coronal sections. Coronal sections were cut serially at 10 μ thickness with rotary microtome. Sections were deparaffinized in xylene (two changes of 10 min each). The sections were hydrated to water through a series of alcohol i.e., 100%, 90%, 70%, 50%, and 30%, 5 min in each grade. The sections were stained in Cresyl violet working solution for 2 min quickly washed in DW and air dried for 45 min in a dust free chamber. The sections were dehydrated in n-butanol, cleared in xylene and mounted in Distretricresyl phosphate xylene (DPX). The brain of control and all the experimental mice were studied with the help of stereotaxic atlas of mice brain and neuronal counting were carried out.

RESULT

MDA estimation
The mean value of MDA content in brain homogenate showed significance (**a, *P* < 0.001) increase in the MDA activity [Figure 1] in group II, and that of group III a significant decrease (**b, *P* < 0.001) was found, when compared to the control. Group IV (**c, *P* < 0.001) shows the MDA concentration near to group I.
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Cresyl violet staining and cell counting
Sections of (Cornu Ammonis area 1) CA1, (Cornu Ammonis area 2) CA3, and (Dentate gyrus) DG region of group I showed optimal sized, pyramidal shaped neuronal cells with a continuous cell membrane [Figure 3]. The cells were closely grouped together to form a band. Group II show extensive degeneration, which appeared to have bursted and lost their shapes, the number of viable cells have been decreased significantly \( P < 0.05 \) [Figure 2] whereas, significant \( P < 0.05 \) increase in the viable cell number was found in group III animals. The morphological structure and number of cells of group IV were found comparable to group I.

DISCUSSION

A number of studies and animal experiments indicate that the involvement of free radicals, LPO, and altered antioxidant defense systems are thought to play a key role in the toxic effects of F.[3] In our study, we found that MDA content in brain tissue was increased, this could be associated with peroxidation of membrane phospholipids and thus accumulation of MDA. F may cross the blood brain barrier, accumulate in mice hippocampal neuron cell bodies and initiates cascade of reactions which increases the oxidative stress that causes an increase in MDA content of brain tissue, which is an end product of LPO.[6] On the other hand, a distinctive decrease in MDA level was observed after Cur treatment, compared to group treated with F.[7] This can be correlated with the fact that this phytochemical is capable of quenching oxygen free radicals such as superoxide anions and hydroxyl radicals, which are important for the initiation of LPO.[8]

At cytological level, neurodegenerative changes have been reported from our laboratory.[9] Although earlier studies on F intoxication and mental disability were observed in fluorosis patients, but these studies were not fully supported with experimental evidences. Recent studies from our laboratory[10,11] reported first time that F causes severe neurodegenerative changes in brain particularly in the hippocampus and cerebral cortex. Treatment with F (120 ppm) for 1 month cause significant degeneration in cells of the hippocampus of mice brain.[12] These neurodegenerative changes are observed as dark cells, pyknotic cell, 5% cells showed reduced size and few cells showed small or shrunken nucleus in CA1, CA3, and DG region of the hippocampus [Figure 3] and in these regions, various degenerative changes were observed. Even the number of neuronal cells counted in a specific region was less in F group animals as compared to control group [Figure 2]. In group III decrease in neurodegenerative effects were observed. It is evident by presence of fewer dark cells, cells with distorted shape etc., A significant increase in viable neuronal cell count was found when the results were compared with the group II.[13] Cresyl violet staining of Cur group clearly showed control pattern of cellular morphology and distribution. Thus Cur protects the neuronal cell bodies in hippocampal area, offering neuroprotection.[24]

CONCLUSION

Our study thus demonstrate that daily single dose of 120 ppm F result in highly significant increases in the LPO as well as neurodegenerative changes in neuron cell bodies of selected hippocampal regions. Supplementation with curcumin significantly reduce the toxic effect of F to near normal level.
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by augmenting the antioxidant defense through its scavenging property and provide an evidence of having therapeutic role against oxidative stress mediated neurodegeneration.

AKNOWLEDGMENT

The authors are thankful to the University Grants Commission, New Delhi for providing a research grant under the UGC-SAP program. Financial assistance was provided to Prof. Maheep Bhatnagar (Coordinator-SAP) under research thrust area–Neurobiology, Gene, and Genomics.

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Figure 3: Photomicrographs demonstrating (Cornu Ammonis) CA1, CA3 and Dentate gyrus region of Hippocampus showing Cresyl violet staining in Control, 120 ppm fluoride (F), Curcumin (Cur) and F + Cur treated group
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Source of Support: Nil, Conflict of Interest: None declared.